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(71) Applicant: **Molecular Diagnostics, Inc., 400 Morgan Lane, West Haven, CT.06516 (US)**

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Bulletin 85/38

(72) Inventor: **Dattagupta, Nanibhushan, 470 Prospect St., New Haven, CT 06511 (US)**
Inventor: **Knowles, William J., 101 Sleeping Giant Dr., Hamden, CT 06518 (US)**
Inventor: **Marchesi, Vincent T., 179 Prospect Ave, Guilford, CT 06437 (US)**
Inventor: **Crothers, Donald M., Surrey Drive, Northford, CT 06472 (US)**

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(74) Representative: **Adrian, Albert; Dr. et al; c/o BAYER AG Konzernverwaltung RP Patentabteilung, D-5090 Leverkusen 1, Bayerwerk (DE)**

(54) **Nucleic acid-protein conjugate.**

(57) A protein is covalently coupled to at least one end of a nucleic acid which carries several labels. In an assay the protein will specifically recognize some component of a test system; in an immunoassay the protein can be Protein A which will recognize the Fc portion of IgG which is bound to an unknown antigen if present in the test sample.

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⑰ Nucleic acid-protein conjugate.

⑰ A protein is covalently coupled to at least one end of a nucleic acid which carries several labels. In an assay the protein will specifically recognize some component of a test system; in an immunoassay the protein can be Protein A which will recognize the Fc portion of IgG which is bound to an unknown antigen if present in the test sample.

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NUCLEIC ACID-PROTEIN CONJUGATE

The present invention relates to the labeling of proteins by covalently coupling them with labeled nucleic acids and the use of these labeled materials in immunoassays for diagnostic purposes.

5 Immunological reagents, e.g., antibodies, are protein molecules that are produced by the body in response to the introduction of a foreign substance known 10 as an antigen. Specific antibodies bind to specific antigens creating antigen-antibody complexes. Since antibodies bind only to their complementary antigens, they can be used to detect the presence of specific antigens in different biological samples. Such 15 immunological methods of detection and diagnosis of certain disease states have revolutionized the biomedical sciences. Antibodies are now routinely used to measure the presence of small quantities of special molecules (e.g., proteins, hormones, drugs) in blood, urine, and 20 other body fluids.

One method commonly employed to detect the specific interaction between an antigen and its corresponding antibody is the radioimmunoassay (RIA), 25 discovered by Yalow and Berson for the detection of insulin. (R.S. Yalow and S.A. Berson; J. Clin. Invest. 1960: 39: 1157-1175). RIA relies on the fact that the antigen being assayed is first labeled with a radioisotope and is then quantitated by standard means of 30 counting radioisotopes. Each kind of radiolabeling has limitations, for example, sensitivity of detection, isotope half-life, hazard to the operator and the problem of waste disposal. The expense of the equipment necessary to count radioisotopes is also considerable. 35 Therefore, it is highly desirable that immuno-labeling be accomplished without resort to radioactivity.

Although there are several nonradioactive immunoassay methods in practice, the level of sensitivity which can be reached by RIA has not been duplicated.

5 One type of assay, for example, is carried out by chemically treating antibodies with fluorescent compounds so that the location of the fluorescent-labeled antibody can be identified with the use of a special fluorescent microscope. The use of fluorescent-labeled 10 antibodies is a widely used procedure with many applications, but this technique also has certain limitations, among them the fact that antibodies labeled by existing methods are not sensitive enough to detect small quantities of antigenic material. To increase 15 sensitivity, polylysine has been used to couple fluorophores but high background problems are created because polylysine binds nonspecifically to many types of compounds.

20 Other types of labeling reagents include protein-bacteriophage conjugates, stable free radicals, electron dense compounds, luminescent compounds and enzymes. All of these labeling agents have certain disadvantages. Certain reagents, for example, enzymes, 25 are difficult to store. Certain labeling reagents are not sensitive enough unless they are present in multiple copies. If multiple copies of label are directly coupled to the reactants, as in the case of fluorescent label, the antibody may lose its capacity to bind to antigen, 30 i.e., become less specific. [Review: A.H.W.M. Schuurs & B.K. Van Weemen, *clin. chim. Acta* 81, (1977) 1-40; D.S. Smith et al. *Ann clin. Biochem.* 18 (1981) 253-274] Therefore, it is highly desirable to label immunological 35 reagents indirectly, to avoid such a decrease in specificity.

European Patent Application No. 84 107 624.3

discloses a photochemical method of labeling nucleic acids. This
5 invention, preferably employed for detection purposes in
hybridization assays, can be used in immunological
assays. The labeling employs a photoreactive
furocoumarin or phenanthridium compound to link the
nucleic acid to a label which can be assayed in a
10 conventional manner. The end product is thus a labeled
nucleic acid probe comprising (a) a nucleic acid
component, (b) a furocoumarin or phenanthridium compound
photo-chemically linked to the nucleic acid component,
and (c) a label chemically linked to (b). For coupling
15 to DNA, aminomethyl trioxsalen, aminomethyl angelicin and
amino alkyl ethidium or methidium azides are the
preferred linking reagents. The nucleic acid component
can be single or double stranded DNA or RNA or fragments
thereof. The label can be anything which can be assayed
20 in a known manner, for example, a hapten such as biotin,
an enzyme such as β -galactosidase, alkaline phosphatase,
horse radish peroxidase, papain or a phycobiliprotein.

It is accordingly an object of the present
invention to provide a highly convenient and sensitive
25 immunoassay which is nonradioactive.

It is another object of the invention to
provide a means of labeling immunological reagents with
multiple labels, i.e., 20 or more labels per protein
molecule, resulting in an immunoassay which is highly
30 sensitive, e.g., as sensitive as radioimmunoassay.

It is another object of the invention to label
the immunological reagent in such way that there is no
interference with the immunological reaction.

These and other objects and advantages are
35 realized in accordance with the present invention
pursuant to which there is provided a protein covalently

coupled to one end of a nucleic acid. The nucleic acid is the label carrier. Since polynucleotides can be covalently modified to carry large numbers of fluorescent and immunogenic molecules, they serve as a matrix for 5 multiple labeling. A nucleic acid molecule is heavily labeled with, for example, a fluorescent molecule or hapten. Then, the labeled nucleic acid is covalently linked to a protein, for example, protein A.

10 The covalently coupled protein-nucleic acid can carry a large number of labels, on the nucleic acid, advantageously applied before the covalent coupling and before use of the coupled material in an immunoassay.

15 The labels may be of any known type, e.g., a fluorophore, an antigen, a hapten, an enzyme, a substrate of an enzyme, a dye stuff, a ligand capable for binding a desired protein, a radioactive isotope, a cofactor or a carbohydrate.

20 In use, the protein moiety of the coupled material is specific for a ligand or some other reagent such as another protein, a specific nucleotide sequence of a nucleic acid, as, for example, repressor proteins or a 25 hapten, which latter can be detected by a secondary reaction with anti-hapten antibodies. Through this specificity, in effect, the labels become "attached" to the site to which the protein specifically binds.

30 The invention also extends to the process of conducting an assay for an antigen, as determined by the presence of a label, wherein an antigen, if contained in a sample, is complexed with an antibody. The antigen-antibody complex is then bound to a labeled protein through a link between the antibody and the 35 labeled protein. This immunoassay is an improvement over prior immunoassays in that the nucleic acid, which is

covalently coupled at one end to a protein, carries the label to be detected by the assay.

The invention also extends to a
5 radioimmunoassay procedure wherein an antigen which may
be present in a sample is complexed with an antibody
specific for said antigen, the complex is bound to a
covalently coupled protein-nucleic acid, as hereinabove
described, and the nucleic acid portion is thereafter
10 radioactively labeled and the label assayed.

Describing the invention in greater detail, the
protein component can be of many diverse kinds, provided
it is site specific. Particularly suitable materials
15 include immunoglobulins, protein A, and the like.
Protein A is a 40,000 Dalton single chain polypeptide
isolated from Staphylococcus aureus. In some strains of
S. aureus, it is covalently linked to the peptidoglycan
part of the cell wall. Protein A binds to several IgG
20 classes of antibodies through Fc-binding regions. In
this case, the antibodies are specific for the antigen to
be detected in the test sample. Protein A is very stable
to heat and denaturing agents and can be renatured
following denaturing conditions. [Review: J.W. Goding:
25 Use of Staphylococcal Protein A as an immunological
reagent, J. Imm. Methods 20 241-253 (1978)]. The binding
of Protein A to IgG does not affect the antigen binding
site of the IgG molecule.

30 Alternatively, the protein moiety can be an
antibody such as IgG and will thus be specific for some
antigen in the test sample. The antigen can be soluble,
intrinsically or chemically bound to a cell surface or
particulate substance. In general, all antigens can be
35 detected using the NAPA reagents.

The diagnostic antigens which may be detected in an unknown sample include lymphocyte markers, tumor specific antigens, tumor cell markers, membrane receptors, microbial antigens, antigens representative of blood types and histocompatibility types, and antigens in serum, urine, etc., which are either elevated or depressed in various disease states.

The antibody has the specificity for a particular antigen. The antibody can be either monoclonal or polyclonal and can be one or more types, e.g., IgG, IgM, and the like. The antibody can be produced in vivo by a number of species or in vitro by cell culture or recombinant DNA technology.

The nucleic acid can be single or double stranded DNA, RNA, DNA-RNA hybrid, oligoribonucleotide or oligodeoxyribonucleotide.

The protein can be covalently coupled to the nucleic acid in various ways. In one procedure, one end of the nucleic acid is modified as with a terminal transferase so as to establish a ribonucleotide end, if not already present. The ribonucleotide end can be oxidized with periodate to form terminal aldehyde groups which can undergo a Schiff's base reaction with an amino-group of the protein, followed by hydrogenation to form a stable aminomethyl linkage between the protein and nucleic acid. If a nucleic acid is desired which carries a -NH_2 moiety, the terminal aldehyd group is reacted with a diamine.

A second procedure may be an enzymatic coupling of a modified nucleotide with a terminal transferase to one end of the nucleic acid. This nucleotide has a modified structure on its heterocyclic ring, i.e. the purine or pyrimidine ring carries a SH or Hg radical.

The label can be applied to the nucleic acid moiety before or after coupling to the protein and even at a much later stage in the assay. Because of its presence on the nucleic acid, the label will not interfere with the reactions of the protein moiety even though many labels may be present per protein moiety, e.g., 20 to 100 or even more. It has been previously shown that if certain proteins are directly labeled, as few as 6 labels are enough to interfere with the reaction.

The label can be attached to the nucleic acid as described in detail in European Patent Application No. 10 84 107 624.3. Representative labels include fluorophores, for example, fluorescein, texas red, rhodamine or phycoerythrine; a hapten or antigen which can serve as a target for secondary labeled antibody; biotin which can be detected by labeled avidin or antibiotin antibody; an enzyme such as horseradish peroxidase or β -galactosidase assayed in a conventional manner; a cofactor for luminescent assay such as FAD or β -galactosidase; a fluorescent modification reagent which effects energy transfer or quenching; a radioisotope; or a carbo-hydrate.

As noted, the protein moiety should be site specific, as for example, to the Fc portion of an antibody such as IgG.

If the label of the NAPA complex is a fluorophore, the presence of the antigen, which reacts with the IgG-NAPA complex, can be determined by direct fluoroimmunoassay. If the label is a hapten, secondary amplification is necessary wherein an IgG molecule specific for the hapten is used. If the label is an enzyme, an enzyme assay can be used by which the amount of bound enzyme can be determined by the enzyme's catalytic reaction of its substrate in a known manner. If the label is a radioisotope, radioimmunoassay is performed in a conventional manner. In the case of a secondary amplification with an IgG specific for the

label of the primary NAPA complex, read-cut can be accomplished by addition of a second labeled NAPA complex carrying a different label.

5 The covalently coupled protein-nucleic acid can be used in immunoassays in a conventional manner, the labels either being present initially or being added at any stage prior to final assay.

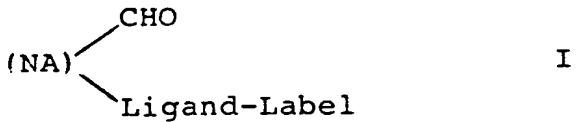
10 The invention will now be described in further detail with reference to the accompanying drawings wherein:

15 Fig. 1 is a schematic flowsheet of a process for coupling Protein A or IgG to a nucleic acid in accordance with one embodiment of the invention; and

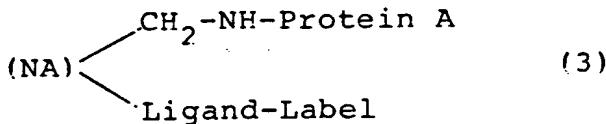
20 Fig. 2 is a schematic flowsheet of a method of using a coupled product of Fig. 1 in an assay.

25 Referring now more particularly to the drawings, in Fig. 1 a nucleic acid containing a terminal ribose residue (NA_r) is covalently linked to a ligand which binds nucleic acids, by photochemical methods (hv). If the NA_r is DNA, the NA_r is prepared from TdT-ATP (terminal deoxynucleotidyl transferase-adenosine triphosphate). If the NA_r is RNA, it is used as such. The covalent adduct, (NA_r)_{ligand}, is formed. The photo-chemical method employs, for example, a photoreactive ethidium azide or psoralen derivative as described in European Patent Application No. 84 107 624.3. If aminomethyl psoralens such as aminomethyl-trioxsalen (AMT) or aminomethyl dimethyl-angelicin (AMA) are used, secondary labeling with fluorescein isothiocyanate (FITC) is possible. The adduct, for example, (NA_r)_{ligand-label}, is formed which, by oxidation with

periodate (NaIO_4), forms aldehyde groups

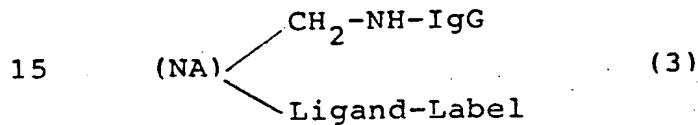


5 This complex, (1), reacts with Protein A and is reduced at (2) with NaBH_4 to form the NAPA complex



10 If, instead of Protein A the complex (1) reacts with IgG, and is reduced at (2),

NA-IgG forms



In place of the fluorescein label, other labels such as haptens, enzymes, cofactors, antigens, 20 radioactive isotopes, carbohydrates and other fluorophores, can be similarly employed.

Referring more particularly to Fig. 2, Protein A is covalently coupled to the ends of a nucleic acid which carries several physically or chemically detectable labels. 25 The label (*) can be a fluorophore if direct fluorescence measurement is desired. For preparation of an enzyme based immunoassay the label can be an enzyme or a substrate of an enzyme. Regarding the coupling of an enzyme such as Horse radish peroxidase about 10 - 100 copies of this enzyme can be coupled. If radioactive measurement is desired the nucleic acid is labeled with a suitable radioactive isotope. The nucleic acid can be a single or double stranded DNA, RNA, DNA-RNA hybrid, oligoribonucleotide or deoxyribonucleotide.

35 The labelled Protein A-DNA adduct (NAPA) will

react with an IgG or an IgG-antigen complex. The complex can then be assayed in conventional manner.

5 If one desires further amplification a hapten or a suitable antigen is used as label. Then the foregoing complex is reacted with an antibody specific for the labels, leaving the Fc fragments of the antibodies free to bind additional quantities of NAPA (Reagent I). This can then be assayed as before.

10 The invention will be further described in the following illustrative examples wherein all parts are by weight unless otherwise expressed.

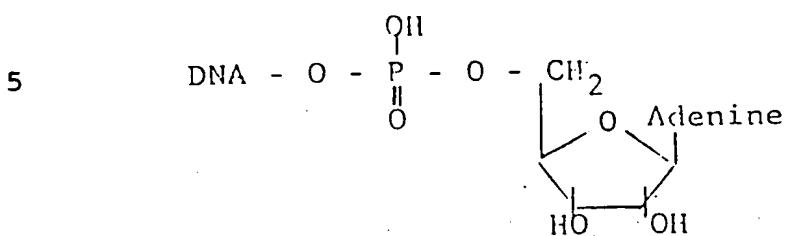
Example 1

Terminal transferase reaction to couple ribose residue at the end of DNA.

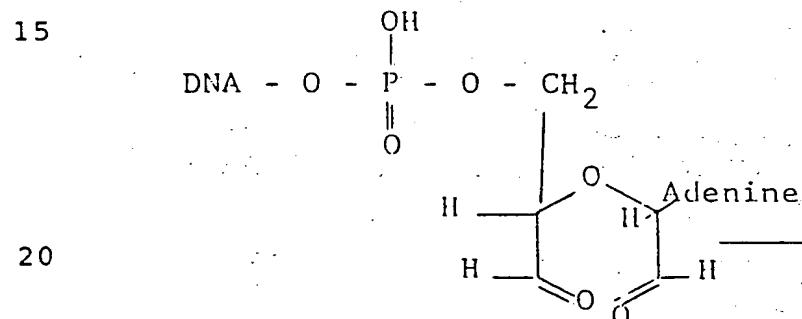
15 Hae III digest of QX 174 RF or the product of Example 3 is dialyzed against potassium cacodylate buffer (pH 7.2; 200 mM) and the concentration is adjusted to 10^{-4} M in base pairs. To the DNA solution (100 μ l), 5 μ l 2 mM dithiothreitol 1 μ l 10 mM ATP mixed with 14 C labeled 20 ATP and 10 μ l 10 mM cobalt chloride are added. The mixture is incubated at 37°C for 5 minutes and then chilled in ice for 10 minutes. 15 units of terminal deoxynucleotidyl transferase is added and the mixture incubated at 15°C for 60 minutes. After the reaction, 25 the enzyme is removed by phenol extraction and the unreacted nucleotides are removed by dialysis or passage through a Sephadex G-50 column. The yield is calculated from the absorbance to CPM ratio. Assuming uniform reaction, the yield is about 10-20 ATP residues per 3' 30 end of the DNA. (For RNA-DNA hybrid or double stranded RNA, this step of TdT reaction is not required for further reaction.)

Example 2

Oxidation and coupling of proteins.



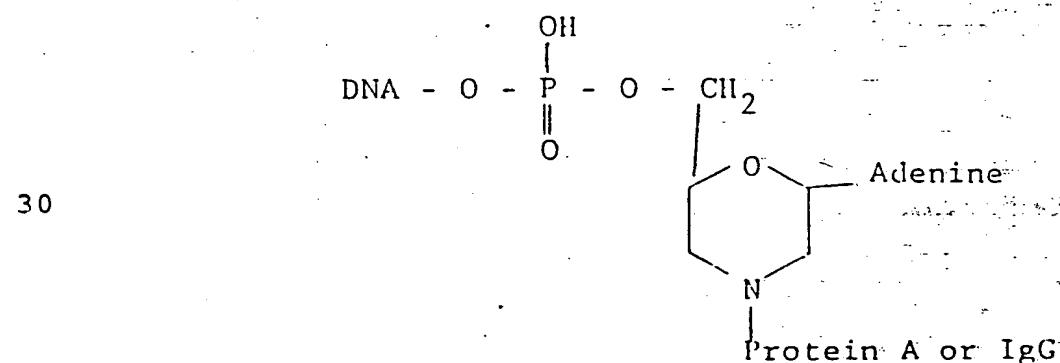
10 0.1 M sodium acetate
pH 5; 0.1 volume 1 M
sodium periodate



20

- 1) 0.05 M Sodium Acetate, pH 9. 1 M NaCl
- 2) Protein A or IgG
- 3) Borohydride reduction

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35 The oxidation of the ribose end of the product of Example 1 is effected with sodium periodate at room temperature using 0.1 M sodium acetate buffer pH 5 and 0.1 volume 1 M sodium

periodate. The resulting solution is dialyzed to remove excess periodate against 1 M NaCl, 0.05 M sodium acetate pH 9. Then Protein A or IgG is added and the solution is reduced by adding Na-borohydride.

5

The reaction can be carried out between 0-50°C and in ionic strength up to 1 M NaCl.

10 Example 3

Covalent reaction between double stranded nucleic acids and aminomethyl trioxsalen:

Hae III restriction enzyme digest of QX174 RF double stranded DNA treated as described in Example 1 or the product of Example 2 is dissolved in or dialyzed against Tris-EDTA buffer (TE) (10 mM trishydroxymethyl aminomethane, 1 mM EDTA; pH 7.1 adjusted with HCl). The concentration is adjusted to 1.5×10^{-4} M/l in base pairs. 4'- aminomethyl trioxsalen (AMT) dissolved in TE is added so that the ratio between the ligand to base pairs is 0.1. The mixture is flushed with nitrogen gas for 15 seconds and irradiated at 360 nm radiation for 60 minutes. This causes the trioxsalen residues to bind the DNA covalently. After the reaction, the mixture is precipitated with ethanol after adding sodium acetate to raise the ionic strength. The precipitation also leaves any unreacted trioxsalen in the supernatant. By using tritiated trioxsalen, the relative amount of coupling can be estimated. (Exactly identical conditions can be employed for double stranded RNA and RNA-DNA hybrids).

Example 4

Labeling of products of Example 2 and 3:

35 The products of Examples 2 and 3 have free primary NH_2 residues from AMT and/or protein. They are

reacted with biotin and/or fluorescein or rhodamine. An example is provided with fluorescein. A fluorescein isothiocyanate (FITC) solution is prepared by dissolving 5 mg of the solid in 2 ml ethanol. Products of Examples 5 and 3 are dialyzed again or dissolved in 0.1 M sodium bicarbonate buffer (pH 8-9). Then they are mixed with ethanolic solution of FITC in 15:1 volume ratio with equal weight concentration. The reaction is allowed to proceed for one hour. The product is purified on 10 Sephadex G50 column. The excluded volume fractions contain the product. Labeling with biotin is done by using N-hydroxysuccinimido biotin as described in European Patent Application No. 84 107 624.3.

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Example 5

Poly allylamino uridine phosphate (produced by 20 enzymatic polymerization of allylamino uridine triphosphate in conventional manner) is reacted with fluorescein, rhodamine or biotin, as in Example 4, then reacted with protein as in Example 2.

The order of reactions can be changed, e.g., 25 DNA can be reacted first with AMT, then ribose residue can be added by TdT, all AMT amine residues can be reacted with labels, and then coupling via redox reaction with proteins can be carried out.

30 Example 6For detection of tumor-specific antigen-carcino-embryonic antigen (CEA).

CEA is a glycoprotein and quantitation of serum 35 CEA has been shown to have a prognostic and potentially diagnostic value in patients with ovarian, breast and colonic carcinoma.

A serum sample (0.1-1 ml) is incubated in a support (e.g., nitrocellulose) so as to allow coupling of the CEA to such a support. The coupling may be non-specific, e.g., adsorption of protein to plastic or 5 may be specific, e.g., immobilization of CEA to a support using an antibody or Fab fragment specific for CEA, the antibody or fragment having been previously immobilized on this support.

10 The immobilized CEA is then incubated with a polyclonal antibody (selected for its antigenic specificity and Protein A binding characteristics and titrated to give maximum specific binding to the antigen) for 30 minutes at room temperature. Non-bound protein is removed by washing three times in phosphate buffered 15 saline (20 mM sodium phosphate, 130 mM NaCl), containing 0.05% Tween-20 (PBS-Tw).

20 The fluorescein-labeled DNA-Protein A complex is incubated with the antibody-antigen complex for 30 minutes at room temperature and washed three times with PBS-Tw to remove non-bound NAPA complex.

The label is then assayed by fluorescent microscopy.

25 Example 7

Analysis of histocompatibility types.

20 Lymphocytes are separated from whole blood (1 ml) using a Ficoll hypaque gradient. Lymphocytes are incubated in a battery of antisera, each antiserum specific for a particular histocompatibility (HLA) 30 sub-type. Nonbound antisera are removed by differential centrifugation and the cells washed three times using PBS. The washed lymphocytes are incubated with the fluorescein-labeled Protein A-DNA complexes. Non-bound 35 fluorescein-labeled Protein A-DNA complexes are removed by differential centrifugation. The lymphocytes are then analyzed by fluorescent microscopy.

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It is understood that the specification and examples are illustrative but not limitative of the present invention and that other embodiments within the spirit and scope of the invention will suggest themselves to those skilled in the art.

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MD 210

WHAT IS CLAIMED IS:

1. A protein covalently coupled to one end of a nucleic acid.

5 2. A coupled protein according to claim 1, characterized in that the nucleic acid is single or double stranded and the said nucleic acid is a deoxy-ribonucleic acid or a ribonucleic acid or an oligodeoxy-ribonucleotide or an oligoribonucleotide or a hybrid thereof.

10 3. A coupled protein according to claim 1 or 2, characterized in that the nucleic acid carries a Hg, SH or NH₂ functional group.

4. A coupled protein according to claims 1 - 3, characterized in that the protein is one which has an affinity for

15 a) another protein,
b) a specific nucleotide sequence of a nucleic acid,
c) a hapten, or
d) a ligand.

20 5. A coupled protein according to claims 1 - 4, characterized in that the nucleic acid carries at least one physically or chemically detectable label.

25 6. A coupled protein according to claims 1 - 5, characterized in that the label can be a fluorophore, a dye stuff, an antigen, a ligand, an enzyme, a substrate of an enzyme, a cofactor, a radioactive isotope, a carbohydrate or a combination thereof.

7. A method for determination an analyte wherein the analyte is complexed with a specific binding compound, characterized in that the specific binding compound is linked to one end of a labeled nucleic acid.

5 8. A method for determination an analyte according to claim 7, characterized in that the specific binding compound is a protein which has an affinity for

10 a) another protein,
b) a specific nucleotide sequence of a nucleic acid,
c) a hapten, or
d) a ligand.

15 9. A method for preparation a protein which is covalently coupled to one end of a nucleic acid, characterized in that the nucleic acid is subjected to chemical or enzymatical treatment to introduce a functional group at one end capable for binding protein.

20 10. A method for preparation of a protein which is covalently coupled to one end of a nucleic acid, characterized in that the nucleic acid carries a ribose moiety at one end which is oxidized to form pendent aldehyde groups which are capable for binding proteins via a Schiff-Base which is reduced if necessary.

25 11. A method for preparation of a protein which is covalently coupled to one end of a nucleic acid according to claim 9 and 10, characterized in that the nucleic acid is labeled with at least one physically or chemically detectable label.

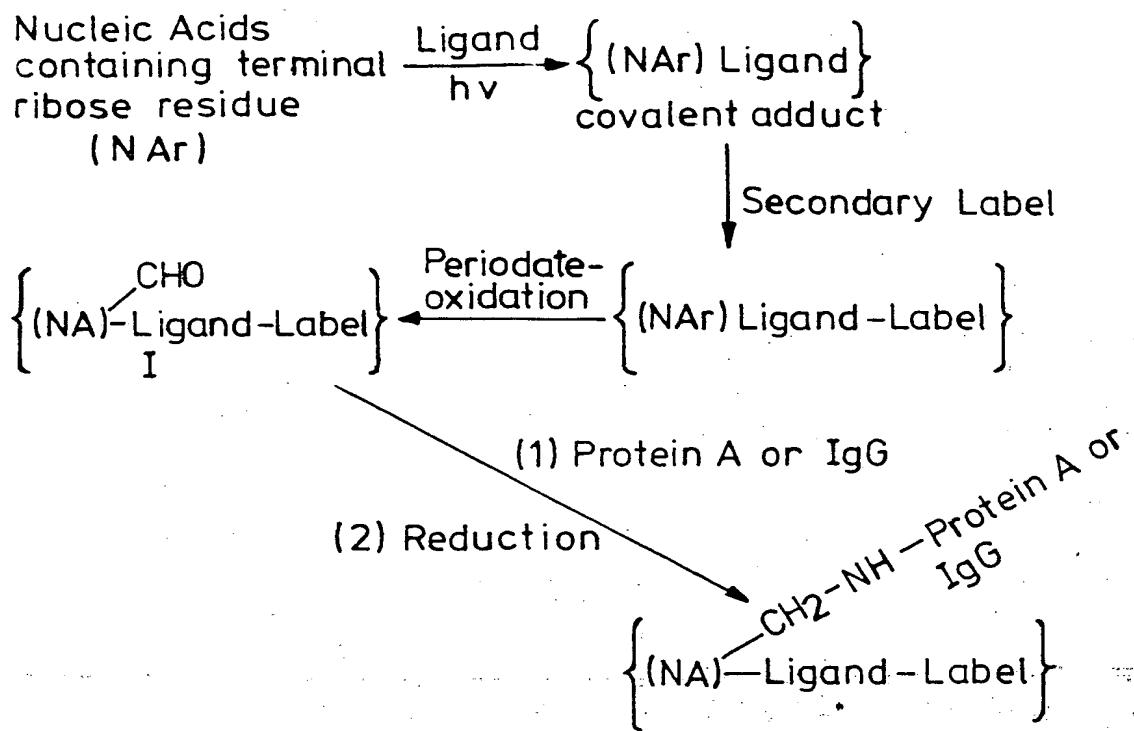


FIG. 1

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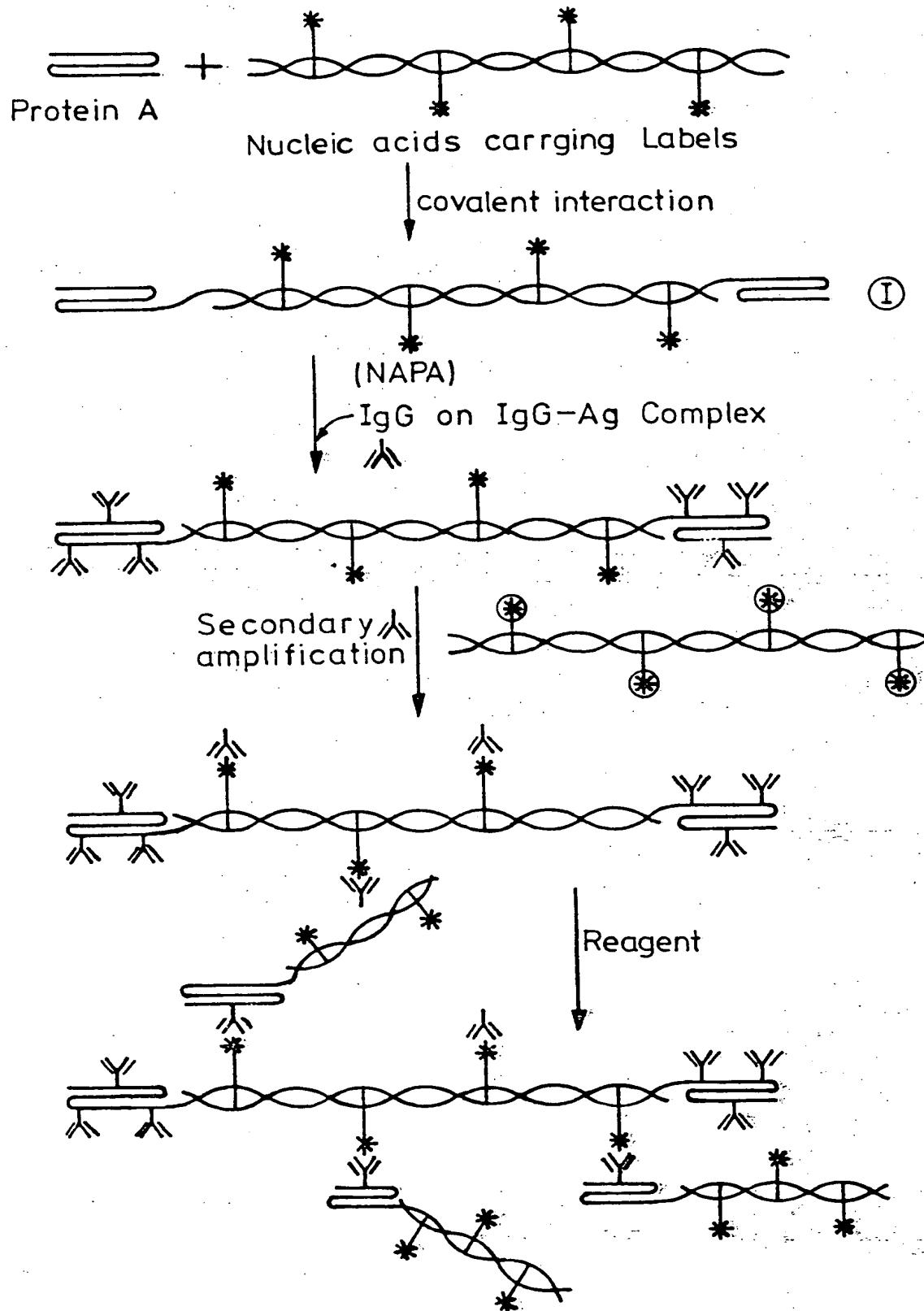


FIG. 2



European Patent
Office

EUROPEAN SEARCH REPORT

0154884

Application Number

EP 85 10 2130

DOCUMENTS CONSIDERED TO BE RELEVANT

Category	Citation of document with indication, where appropriate, of relevant passages	Relevant to claim	CLASSIFICATION OF THE APPLICATION (Int. Cl. 4)
X	GB-A-2 019 408 (INSTITUT PASTEUR) * Page 4, lines 44-51; page 5, line 22 - page 6, line 3 * ---	1,2,4-9 ,11	G 01 N 33/532 C 12 Q 1/68 G 01 N 33/574 G 01 N 33/50
X	US-A-4 358 535 (S. FALKOW et al.) * Column 1, line 64 - column 2, line 17; column 2, lines 43-44; column 3, line 28 - column 4, line 18 * ---	1,2,4-9 ,11	
X,P	WO-A-8 403 520 (A.D.B. MALCOLM et al.) * Page 3, lines 22-32; page 5, line 21 - page 6, line 11; page 8, lines 23-33; page 12, line 23 - page 13, line 10; page 24, line 1 - page 25, line 19 * ---	1,2,4-9 ,11	
X,P	EP-A-0 131 830 (MOLECULAR DIAGNOSTICS) * Page 8, line 19 - page 9, line 23; page 16, example 4; page 22, lines 2-5, 15-18 * ---	1,2,4-9 ,11	
X,P	GB-A-2 125 964 (CETUS MADISON CORP.) * Page 1, lines 76-95; page 3, lines 81-83, 97-112; page 10, lines 13-30 * ---	1,2,4-9 ,11	TECHNICAL FIELDS SEARCHED (Int. Cl. 4)
X,P	US-A-4 587 044 (P.S. MILLER et al.) * Column 1, lines 42-49; column 2, line 60 - column 3, line 60; column 21, line 64 - column 24, line 14 * ---	1,2,4-9 ,11 -/-	G 01 N C 12 Q

The present search report has been drawn up for all claims.

Place of search	Date of completion of the search	Examiner
THE HAGUE	15-09-1988	VAN BOHEMEN C.G.

CATEGORY OF CITED DOCUMENTS

X : particularly relevant if taken alone
Y : particularly relevant if combined with another document of the same category
A : technological background
O : non-written disclosure
P : intermediate document

T : theory or principle underlying the invention.
E : earlier patent document, but published on, or after the filing date
D : document cited in the application
L : document cited for other reasons
& : member of the same patent family, corresponding document



DOCUMENTS CONSIDERED TO BE RELEVANT

Category	Citation of document with indication, where appropriate, of relevant passages	Relevant to claim	CLASSIFICATION OF THE APPLICATION (Int. Cl. 4)
X	CHEMICAL ABSTRACTS, vol. 88, 1978, page 219, no. 132899b, Columbus, Ohio, US; M. KROEGER et al.: "Nucleophilic substitution at C-2 of S-alkylated 2-thiocytidines by cysteine and lysine. A new method for specific covalent linking of peptides to nucleic acids" & BIOORG. CHEM. 1977, 6(4), 431-41 * Abstract * ---	1-3	
X	CHEMICAL ABSTRACTS, vol. 93, 1980, page 399, no. 64767h, Columbus, Ohio, US; Y.-C. TSE et al.: "Covalent bonds between protein and DNA. Formation of phosphotyrosine linkage between certain DNA topoisomerases and DNA" & J. BIOL. CHEM. 1980, 255(12), 5560-5 * Abstract * ---	1,2	
X	CHEMICAL ABSTRACTS, vol. 98, 1983, page 229, no. 157399h, Columbus, Ohio, US; S. CHENG et al.: "A versatile method for the coupling of protein to DNA: synthesis of a2-macroglobulin-DNA conjugates" & NUCLEIC ACIDS RES. 1983, 11(3), 659-69 * Abstract * -----	1,2	TECHNICAL FIELDS SEARCHED (Int. Cl. 4)

The present search report has been drawn up for all claims

Place of search	Date of completion of the search	Examiner
THE HAGUE	15-09-1988	VAN BOHEMEN C.G.
CATEGORY OF CITED DOCUMENTS		
<p>K : particularly relevant if taken alone Y : particularly relevant if combined with another document of the same category A : technological background O : non-written disclosure P : intermediate document</p> <p>T : theory or principle underlying the invention E : earlier patent document, but published on, or after the filing date D : document cited in the application L : document cited for other reasons & : member of the same patent family, corresponding document</p>		